Research Article



Tegumental Alterations on Raillietina galli, an Intestinal Parasite of Fowl, after Treatment with Millettia taiwaniania Extract

KHOLHRING LALCHHANDAMA*

Department of Life Sciences, Pachhunga University College, Aizawl 796 001, Mizoram, India.

Abstract | *Millettia taiwaniana* is a multipurpose medicinal plant used in the treatment of several diseases. The root bark is used by the Mizo people of India and Myanmar for the treatment of intestinal helminthiasis. The study aims to reveal the anthelmintic effects on a parasitic tapeworm of fowl, *Raillietina galli*, using scanning electron microscopy. The tapeworms were treated *in vitro* with the methanol extract of the root bark. The treated cestodes were fixed, dehydrated, and dried. They were then observed using scanning electron microscopy. There was massive tegumental damage throughout the body surface. Erosion and shrinkage were seen on the scolex and body segments. There were numerous pits due to disintegration of the tegumental layer. Folds and blebs were formed on the large gravid proglottids. *M. taiwaniana* extract exhibits powerful anthelmintic effects similar to those established of standard anthelmintic drugs.

Keywords | Anthelmintic, Millettia taiwaniana, Raillietina galli, Microscopy, Tegument

Received | May 25, 2019; Accepted | July 19, 2019; Published | November 01, 2019

*Correspondence | Kholhring Lalchhandama, Department of Life Sciences, Pachhunga University College, Aizawl 796 001, Mizoram, India; Email: chhandama@pucollege.edu.in

Citation | Lalchhandama K (2019). Tegumental alterations on raillietina galli, an intestinal parasite of fowl, after treatment with millettia taiwaniania extract. Adv. Anim. Vet. Sci. 7(11): 1010-1014.

DOI | http://dx.doi.org/10.17582/journal.aavs/2019/7.11.1010.1014

ISSN (Online) | 2307-8316; ISSN (Print) | 2309-3331

Copyright © 2019 Lalchhandama. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Millettia taiwaniana Hayata is a perennial shrub of family Fabaceae that has gained attention in pharmaceutical investigations. It is widely used in traditional practices in different cultures of Southeast Asia as a medicinal plant for the treatment of clinical conditions including cancer, anaemia and immune diseases (Harrison et al., 2011). The roots and seeds are commonly used as insecticides and as fish poison. As haemotopoietic agent, the seed extract has been used as supplementary medicine for the treatment of blood cancer, and it is also known to be directly anticarcinogenic (Ye et al., 2012). Among the Mizo people of India and Myanmar, the root bark is a potent fish poison and antiparasitic agent (Lalchhandama et al., 2008).

Important bioactive compounds such as chalcones, dihydroflanonol, isoflavonoids, and rotenoids are identified from the seed (Ye et al., 2008; Deyou et al., 2017). 4-Hydroxylonchocarpin and deguelin from the seeds have high antiinflammatory activities by inhibiting synthesis of ni-

tric oxide, activity of inducible NO synthase (iNOS) and expression of *iNOS* gene (Ye et al., 2014). Millepachine has been among the promising lead compounds showing broad-spectrum activities against different cancer cells (Wu et al., 2016; Yang et al., 2018).

Species of *Millettia* are known for their abundant contents of rotenoids. Rotenone, *cis*-12*a*-hydroxyretenone, *cis*-12a-hydroxyrot-2'-enonic acid, and rot-2'-enonic acid have insecticidal properties. Isoflavones, β-sitosterol, oleanolic acid, karanjin, and pachycarin A to C are also isolated (Chen et al., 1999; Lu et al., 1999; Shao et al., 2001). Barbigerone, a rotenoid derivative, is an established anticarcinogen (Qiu et al., 2015). Isoflavonoids such as auriculasin, 6,8-diprenylorobol, erysenegalensein E, isoerysenegalensein E, furowanin A and B, and millewanins G (1) and H (2) from the leaves showed strong antiestrogenic activity (Okamoto et al., 2006).

It was previously reported that *M. taiwaniana* root bark extract had anthelmintic activity by causing dosedependent efficacy on poultry tapeworm (Roy et al., 2008),



and the anthelmintic activity was associated with structural damages and biochemical alterations in the parasites (Lalchhandama et al., 2008). Anthelmintic drugs are best evaluated by studying their effects on the fine structure of the worms. Therefore, this study attempted to assess the effects of the plant extract on the tegument of a parasitic cestode.

MATERIALS AND METHODS

PLANT EXTRACT

M. taiwaniana Hayata was collected from Aizawl, India, and identified (PUC-BOT-M-036) as reported earlier (Roy et al., 2008). The root bark was washed, slice into small pieces, and dried. Extract was done with methanol in a Soxhlet apparatus and concentrated in a vacuum rotary evaporator (Buchi Rotavapor® R-215).

IN VITRO SURVIVAL TEST

Live adult cestodes, *Raillietina galli* Yamaguti, 1935, were recovered from the intestines of fowls (*Gallus gallus domesticus* Linnaeus, 1758). They were collected in neutral phosphate-buffered saline (PBS) maintained at 37±1°C. One group was treated with albendazole (20 mg/ml), the second with *M. taiwaniana* root extract (20 mg/ml dissolved in PBS with 1% DMSO), and the third group was maintained as control in PBS with 1% DMSO. Survival time was recorded, and data were analysed using Student's *t*-test, and the level of significance taken at *p* < 0.05.

SCANNING ELECTRON MICROSCOPY

Plant extract-treated cestode was washed with PBS and then fixed in 4% neutral-buffered formaldehyde at 4°C for 4 hours. 0.1 M Sodium cacodylate (pH 7.2) was used as buffer. Secondary fixation was done with 1% osmium tetroxide (OsO₄) at 4°C for 1 h. The fixative was buffered again with sodium cacodylate. The specimens were dehydrated through increasing concentrations of propanone up to pure propanone. After complete dehydration, they were then treated with tetramethylsilane, Si(CH₃)₄, for 15 minutes and left to dry in an air-drying chamber at 25°C. They were mounted on metal stubs and sputter coated with gold in JFC-1100 (JEOL Ltd., Tokyo, Japan) ion-sputtering chamber. Finally, they were observed under a JSM-6360 scanning electron microscope (JEOL Ltd., Tokyo, Japan) at an electron accelerating voltage of 20 kV.

RESULTS

In vitro anthelmintic efficacy of albendazole and M. tai-waniana extract against R. galli are shown in Table 1. Both the drug and the plant extract showed significant (p < 0.05) dose-dependent anthelmintic activity at all concentrations tested. Normalised against the survival value of control, it

took 23.76 \pm 1.93 h and 4.39 \pm 0.88 h for albendazole to completely kill the parasites at the lowest and highest concentrations respectively. The plant extract took 063.09 \pm 2.05 h and 012.91 \pm 0.88 h at similar concentrations.

Table 1: Efficacy of albendazole and *M. taiwaniana* root bark extract on *R. galli*.

Treatment	Dose (mg/ml)	Normalised survival time (hour) in mean ± SD	t value	t critical value
Control	0	100.00 ± 2.56	NA	NA
Albendazole	1.25	023.76 ± 1.93*	58.32	2.26
	2.5	020.24 ± 0.58*	74.53	2.45
	5	016.30 ± 0.66*	77.66	2.45
	10	012.15 ± 0.61*	81.85	2.45
	20	004.39 ± 0.88*	86.56	2.45
M. taiwaniana extract	1.25	063.09 ± 2.05*	26.17	2.23
	2.5	046.90 ± 1.36*	31.57	2.26
	5	039.48 ± 1.52*	49.87	2.31
	10	022.48 ± 1.09*	68.29	2.36
	20	012.91 ± 0.88*	78.85	2.45

*Significantly different at p < 0.05 against control; NA = not applicable; n = 6.

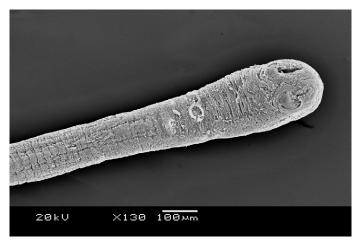


Figure 1: Scanning electron microscopy of the anterior portion of R. *galli* treated with M. *taiwaniana* extract. The terminal bulb is the scolex and the stalk is the neck. Two suckers are seen on the scolex. x 130, scale bar is 100 μ m.

Structural changes on the body of *R. galli* after exposure to *M. taiwaniana* extract was examined by scanning electron microscopy. The plant extract exerted typical anthelmintic effects by causing extensive damage on the tegument throughout the body surface. The effect on the anterior part of the body is shown in Figure 1. There is severe constriction of the tegument with numerous irregular folds. Erosion and scarring are evident throughout. The attachment organs (suckers) are completely damaged and all the spines are sloughed off. There are large pits at the

centre of each sucker indicating heavy erosions. Proglottids (body segments) of the neck region indicate with extensive erosion of the tegument (Figure 2). There are no signs of smooth hair-like microtriches, which are otherwise the surface components of proglottids. Minutes pits and holes are formed at many places. There is an unusual distention on a portion of the mature proglottids (Figure 3). The distended proglottids indicate extensive erosion with pit formations, along with severe shrinkage of the tegument. Under magnification of the enlarged proglottid, distorted and degenerated microtriches are visible (Figure 4). The last gravid proglottid is particularly shrunk and crooked as shown in Figure 5. The tegument looks as if it is forcefully wrung. Shrinkage is associated numerous folds along the length of the body. The tegumental folds are actually due to severe constriction which produced many blisters or blebs on the surface (Figure 6). These blebs represent degenerated microtriches.

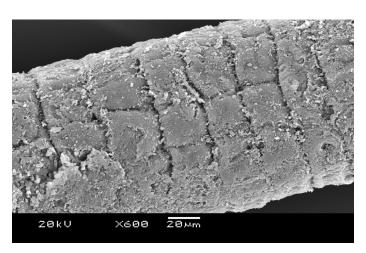


Figure 2: Scanning electron microscopy of the neck of R. *galli* treated with M. *taiwaniana* extract. Body segments or proglottids are heavily eroded. x 600, scale bar is 20 μ m.

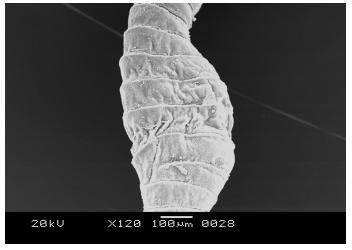


Figure 3: Scanning electron microscopy of mature proglottids of R. *galli* treated with M. *taiwaniana* extract. Abnormal enlargement of the body is evident. x 120, scale bar is 100 μ m.

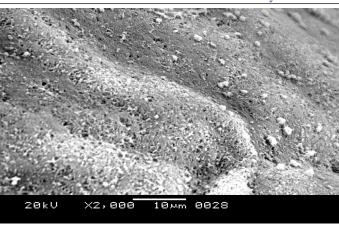


Figure 4: Scanning electron microscopy of a mature proglottids of R. galli treated with M. taiwaniana extract. A fuzzy surface depicts degenerated microtriches. x 2000, scale bar is 10 μ m.

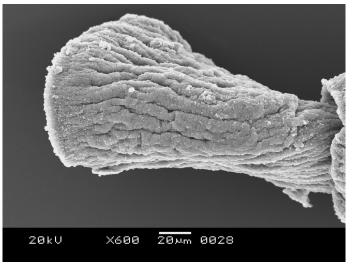


Figure 5: The last gravid proglottid of *R. galli* treated with *M. taiwaniana* extract. Shrinkage with a series of longitudinal folds is visible. x 600, scale bar is 20 μ m.

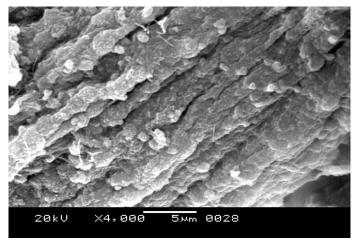


Figure 6: Magnification of the tegument of a gravid proglottid of R. galli treated with M. taiwaniana extract. Tegumental folds are composed of blisters or blebs. x 4000, scale bar is 5 μ m.

REFERENCES

The body surface of cestode is called tegument and is fully covered with hairy microtriches. Without internal digestive and complex nervous system, the tegument acts as a primary absorptive and sensory site, and for this the tegument is the principal target site of anthelmintic drugs. Different anthelmintics are shown to induce surface damages of varying degrees on different cestode. These drugs enter cestode bodies by passive diffusion, and as such, the effects of anthelmintic agents are studied and established upon structural changes they cause on teguments (Rana and Misra-Bhattacharya, 2013; Taman and Azab, 2014).

Albendazole and flubendazole causes rostellar distortion, eruption of swellings or blebs on the tegument, defacement of the microtriches, and increased vesiculation on the human tapeworm, *Echinococcus granulosus* (Elissondo et al., 2006). Albendazole and praziquantel combination caused upon *E. granulosus* and *Mesocestoides corti* resulted in extensive malformation of the suckers, erosion of the tegument and disintegration of microtriches (Li et al., 1996; Markoski et al., 2006). Albendazole also caused severe shrinkage and tegumental collapse in *Raillietina echino-bothrida* (Lalchhandama, 2010).

The fine structure of the body surface of *R. galli* and related species is elucidated (Bâ et al., 1995; Lalchhandama, 2009). *M. taiwaniana* extract showed anthelmintic effects on *R. galli*. Suckers were most conspicuously destroyed on the scolex while the rostellum is hardly affected. in the present study also showed considerable structural damages. Spines were completely removed from the suckers. Erosion on the general tegument was visible throughout the body. Microtriches were reduced to fuzzy debris and the underlying tegumental layers had pits and holes on mature proglottids. These observations indicate that the plant contains important lead molecules for novel anthelmintics and the need for further pharmacological studies.

CONCLUSION

M. taiwaniana root bark extract exhibits anthelmintic activity against parasitic tapeworm of fowl, R. galli. The effects were observed on the tegument of the tapeworms in the form of damages of the suckers, tegumental shrinkage, degeneration of microtriches, surface erosion and formation of blebs.

CONFLICT OF INTEREST

The author declares that there is no conflict of interest.

- •Bâ CT, Sene T, Marchand B (1995). Scanning electron microscope examination of scale-like spines on the rostellum of five Davaineinae (Cestoda, Cyclophyllidea). Parasite. 2: 63–67. https://doi.org/10.1051/parasite/1995021063
- Chen F, Lu J, Chen Q, Guo J, Zhong S, Yan X, Zhu Y (1999).
 Studies on the chemical constituents of thickfruit millettia root (*Millettia pachycarpa*) I. Chinese Tradit. Herbal Drugs. 30: 3–5.
- Deyou T, Marco M, Heydenreich M, Pan F, Gruhonjic A, Fitzpatrick PA, Koch A, Derese S, Pelletier J, Rissanen K, Yenesew A (2017). Isoflavones and rotenoids from the leaves of *Millettia oblata* ssp. *teitensis*. J. Nat. Prod. 80: 2060–2066. https://doi.org/10.1021/acs.jnatprod.7b00255
- Elissondo M, Dopchiz M, Ceballos L, Alvarez L, Bruni SS, Lanusse C, Denegri G (2006). In vitro effects of flubendazole on *Echinococcus granulosus* protoscoleces. Parasitol. Res. 98: 317–323. https://doi.org/10.1007/s00436-005-0026-6
- Harrison JJE, Dankyi E, Kingsford-Adaboh R, Ishida H (2011). In search of new leads: a closer look at the therapeutic potential of the constituents of *Millettia thonningii*, *Millettia pachycarpa* and their structural analogues. Int. J. Pharm. Pharmaceut. Sci. 3: 71–81.
- Lalchhandama K (2009). On the structure of *Raillietina echinobothrida*, the tapeworm of domestic fowl. Sci. Vis. 4: 174–182.
- Lalchhandama K (2010). In vitro effects of albendazole on Raillietina echinobothrida, the cestode of chicken, Gallus domesticus. J. Young. Pharm. 2: 374–378. https://doi. org/10.4103/0975-1483.71630
- Lalchhandama K, Roy B, Dutta B (2008). Effects of *Millettia pachycarpa* on the trace metals and tegumental enzymes of *Raillietina echinobothrida*. Pharmacogn. Mag. 4: 254–261.
- Li S, Wu L, Liu Z, Hu L, Xu P, Xuan Y, Liu Y, Liu X, Fan J (1996). Studies on prophylactic effect of artesunate on schistosomiasis japonica. Chinese Med. J. 109: 848–853.
- Lu J, Zeng J, Kuang Z, Chen F (1999). Studies on the chemical constituents of thickfruit *Millettia* root (*Millettia pachycarpa*) (II). Chinese Tradit. Herbal Drugs. 30: 721–723.
- Markoski MM, Trindade ES, Cabrera G, Laschuk A, Galanti N, Zaha A, Nader HB, Ferreira HB (2006). Praziquantel and albendazole damaging action on in vitro developing *Mesocestoides corti* (Platyhelminthes: Cestoda). Parasitol. Int. 55: 51–61. https://doi.org/10.1016/j.parint.2005.09.005
- Okamoto Y, Suzuki A, Ueda K, Ito C, İtoigawa M, Furukawa H, Nishihara T, Kojima N (2006). Anti-estrogenic activity of prenylated isoflavonoids from *Millettia pachycarpa*: implications for pharmacophores and unique mechanisms. J. Health Sci. 52: 186–191. https://doi.org/10.1248/jhs.52.186
- Qiu N, Cai L, Wang W, Wang G, Cheng X, Xu Q, Wen J, Liu J, Wei Y, Chen L (2015). Barbigerone-in-hydroxypropyl-β-cyclodextrin-liposomal nanoparticle: preparation, characterization and anti-cancer activities. J. Incl. Phenom. Macrocycl. Chem. 82: 505–514. https://doi.org/10.1007/s10847-015-0533-8
- Rana AK, Misra-Bhattacharya S (2013). Current drug targets for helminthic diseases. Parasitol. Res. 112: 1819–1831. https://doi.org/10.1007/s00436-013-3383-6
- Roy B, Lalchhandama K, Dutta B (2008). Scanning electron microscopic observations on the *in vitro* anthelmintic effects of *Millettia pachycarpa* on *Raillietina echinobothrida*. Pharmacogn. Mag. 2008; 4: 20–26.



- Shao W, Zhu Y, Guan S, Zhong S, Chen F (2001). Study on the chemical constituents of thickfruit millettia root (*Millettia pachycarpa*). Nat. Prod. Res. Dev. 13: 1–4.
- Taman A, Azab M (2014). Present-day anthelmintics and perspectives on future new targets. Parasitol. Res. 113: 2425–2433. https://doi.org/10.1007/s00436-014-3969-7
- •Ye H, Chen L, Li Y, Peng A, Fu A, Song H, Tang M, Luo H, Luo Y, Xu Y, Shi J (2008). Preparative isolation and purification of three rotenoids and one isoflavone from the seeds of *Millettia pachycarpa* Benth by high-speed countercurrent chromatography. J. Chromatogr. A. 1178: 101–107. https://doi.org/10.1016/j.chroma.2007.11.060
- Ye H, Fu A, Wu W, Li Y, Wang G, Tang M, Li S, He S, Zhong S, Lai H, Yang J (2012). Cytotoxic and apoptotic effects of constituents from *Millettia pachycarpa* Benth. Fitoterapia. 83: 1402–1408. https://doi.org/10.1016/j.fitote.2012.08.001
- •Ye H, Xie C, Wu W, Xiang M, Liu Z, Li Y, Tang M, Li S, Yang J, Tang H, Chen K (2014). *Millettia pachycarpa* exhibits anti-inflammatory activity through the suppression of LPS-induced NO/iNOS expression. Am. J. Chinese Med. 42: 949–965. https://doi.org/10.1142/S0192415X14500608
- •Wu W, Ma B, Ye H, Wang T, Wang X, Yang J, Wei Y, Zhu J, Chen L (2016). Millepachine, a potential topoisomerase II inhibitor induces apoptosis via activation of NF-kB pathway in ovarian cancer. Oncotarget. 7: 52281–52293. https://doi.org/10.18632/oncotarget.10739
- Yang J, Yan W, Yu Y, Wang Y, Yang T, Xue L, Yuan X, Long C, Liu Z, Chen X, Hu M (2018). The compound millepachine and its derivatives inhibit tubulin polymerization by irreversibly binding to the colchicine-binding site in β-tubulin. J. Biol. Chem. 293: 9461–6472. https://doi.org/10.1074/jbc. RA117.001658

