Research Article



Growth Performance, Morphological and Chemical Characteristics of Red Tilapia Fed Diets Supplemented with Dunaliella salina

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Abstract | The aims of the present study were to investigate growth performances, morphological and chemical body characteristics, blood profiles of Red Tilapia upon dietary Dunaliella salina (D. salina) supplementation. Control and D. salina diets were formulated isonitrogenous and isoenergetic (339.99±0.02 g/kg & 4.5 ± 0.0028 Kcal/g, respectively) and fed during the study. The formulated diets were control diet and D. salina diets supplemented with 33.0, 66.0 and 100% and fed thrice a day to fish (14.37±0.15 g g) for fourteen weeks in 16 tanks containing 30 fish in each. Body weight gain, feed efficiency and morphological and chemical body characteristics of treated *D. salina* fish were determined in addition to changes in flesh color and blood profiles (red blood cells, hematocrit, glucose and total protein) compared to control one. The results indicated that replacement of fish meal with 33.0% D. salina resulted in comparable results with control diet concerning body weight gain, feed efficiency, morphological and chemical body composition and blood profiles. Diet containing 33% of *D. salina* had no significant effect in protein productive value when compared to control diet versus 66 and 100% D. salina diets, which decreased the parameter. Productive value of energy was not differed (P > 0.05) between control and D. salina treated groups. On the other hand, the high levels of D. salina (66.0 & 100%) were mostly decreased all the recorded parameters compared to control and 33.0% D. salina diets except body fat content and flesh color, which were significantly (P < 0.05) improved. In conclusion, replacement of fish meal with 33.0% D. salina could be promising of Red Tilapia growth performances, feed efficiency and chemical characteristics.

Keywords | Dunaliella salina, Red Tilapia, Blood, Growth, Flesh color

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INTRODUCTION

ecent interest has grown for the use of *Dunaliella sali-* R_{na} (D. salina) as a novel feed supplement for nutritional purposes (Senosy et al., 2017; Mohammed, 2018a) or detoxification and chelating of toxic and heavy metals. D. salina grant various health effects to the humans and animals as well. D. salina because of its content of high β-carotene can protect against CNS oxygen toxicity (Bitterman et al., 1994) and gastrointestinal inflammation (Lavy et al., 2003) through anticancer and antioxidant properties (Krinsky, 1988). Moreover, D. salina has immunomodulatory and anti-inflammatory effects (Hemmingson et al., 2006; Abdel-Daim et al., 2015).

D. salina as feed supplement were investigated in different species of fish for its effects on growth performances, fecundity and hematological parameters. Alishahi et al., (2013) investigated the hemato-immunological responses of Heros severus fed diets supplemented with D. salina. They found that D. salina were positively affected the growth, immunological and hematological parameters. In addition, the effects of dietary D. salina extract on the fecundity and lipid content of pond-reared Penaeus japonicus brood-stock were explored (El-Bermawi et al., 2013). The results indicated that D. salina extract might play role in stress tolerance and reproductive performance of beta-carotene content. Recently, biotechnological interests of D.sa



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Table 1: Ingredients (g/kg) and chemical composition of control and Dunaliella salina (DS) formulated diets used during the study

Parameters	Control	DS 33%	DS 66%	DS 100%
Fish powder	169.60	115.60	57.60	0.00
Soybean	280.80	284.70	296.30	310.00
Gluten	100.00	100.00	100.00	100.00
Maize	210.00	200.00	200.00	220.00
Wheat Bran	150.00	134.00	106.60	54.70
D. salina	0.00	70.70	141.50	214.30
Fish oil	64.60	70.00	73.00	76.00
Calcium diphosphate	5.00	5.00	5.00	5.00
Minerals & vitamins	5.00	5.00	5.00	5.00
Sodium choloride	10.00	10.00	10.00	10.00
Limestone	5.00	5.00	5.00	5.00
Total	1000	1000	1000	1000
Chemical composition				
Dry matter, %	93.79	93.49	93.19	92.84
Crude protein, %	34.00	34.00	34.00	34.00
Fat, %	10.48	11.00	11.26	11.56
Crude fiber, %	3.38	3.58	3.76	3.87
Ash,%	7.11	7.15	7.15	7.06
Nitrogen-free extract, %	38.82	37.76	37.01	36.35
Energy,	4.50	4.51	4.50	4.50

Table 2: Concentrations of minerals and vitamins mixtures per kilogram feed

Minerals	Mg/kg
Calcium phosphate dihydrogen	727.7775
Magnesium sulfate	127.50
Calcium carbonate	84.7225
Sodium chloride	60.0
Vitamins	IU or mg/kg
Vitamin A (IU/kg)	2000000
Vitamin D (IU/kg)	300000
Vitamin E (IU/kg)	40000
Vitamin C (mg/kg)	60000
Vitamin K (mg/kg)	1200
Vitamin B1 (mg/kg)	4000
Vitamin B2 (mg/kg)	4000
Vitamin B6 (mg/kg)	2400
Vitamin B12 (mg/kg)	10
Niacin (mg/kg)	24000
Folic acid (mg/kg)	800
Pantothenic acid	5000
Biotin (mg/kg)	200
Antioxidant (mg/kg)	20000
Cobalt (mg/kg)	200
Copper (mg/kg)	1000

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Iron (mg/kg)	4000
Manganese (mg/kg)	15000
Selenium (mg/kg)	20
Zinc (mg/kg)	15000

Table 3: Growth performance of Red Tilapia fed diet supplemented with Dunaliella salina

Parameters	Control	DS 33%	DS 66%	DS 100%
Initial body weight (g/kg)	12.35±0.10	12.49±0.17	12.32±0.21	12.45±0.19
Final body weight (g/kg)	42.51±0.20a	39.46±0.11 ^b	37.52±0.19 ^c	36.87±1.07 ^c
Body weight gain (g/fish)	30.17±0.11 ^a	26.97 ± 0.14^{b}	24.53±1.17°	24.42±1.02°
Body weight, %	244.32±1.33 ^a	216.02±4.01 ^b	204.53±4.88°	196.19±4.16°
Daily body weight gain (g/fish)	0.36 ± 0.00^{a}	0.32 ± 0.002^{b}	0.29 ±0.01°	0.29 ±0.01°
Specific growth rate	1.47±0.005ª	1.37 ± 0.02^{b}	1.32±0.04°	1.29±0.03°
Survival rate, %	85.56±6.94	83.33±6.67	85.56±7.70	80.00±5.77

a,b,c; Values with different superscripts between DS (D. salina) and control groups significantly differed at P < 0.05.

Table 4: Feed efficiency of Red Tilapia fed diet supplemented with Dunaliella salina

Parameters	Control	DS 33%	DS 66%	DS 100%
Average daily feed intake, g	44.33±1.02 ^a	41.31±0.87 ^b	40.07±0.56°	37.69 ± 0.49^{d}
Average dry daily feed intake, g	41.58±0.95 ^a	38.74±0.82 ^b	37.58±0.53°	35.35±0.46 ^d
Daily feed intake, g	2.2± 0.06 ^a	$2.16 \pm 0.05^{\rm b}$	2.19±0.03°	2.08 ± 0.04^{d}
Feed efficiency, %	68.07± 1.8a	65.31± 1.06 ^b	62.88±0.54 ^b	64.79± 2.67 ^b
Feed conversion	1.38 ± 0.04^{b}	1.44±0.02 ^a	1.49±0.01 ^a	1.45 ± 0.06^{a}
Protein				
Protein efficiency	2.0 ± 0.05^{a}	$1.91 \pm 0.03^{\mathrm{ab}}$	1.84 ± 0.01^{b}	1.89± 0.04 ^b
Protein productive value	30.11 ± 0.92^{a}	29.29± 1.72ab	25.45±1.71 ^b	25.30± 0.72 ^b
Energy				
Energy efficiency	15.12 ± 0.40^{a}	14.45 ± 0.23^{ab}	13.88±0.1 ^b	14.24± 0.34 ^b
Energy productive value	23.22 ± 1.40^{a}	23.16±1.48a	22.12±0.27 ^a	23.27±1.32 ^a

a,b,c,d; Values with different superscripts between DS (D. salina) and control groups significantly differed at P < 0.05.

-lina extracts have been arisen and explored. Gallego-Cartagena et al. (2019) investigated the effect of stressful conditions on the carotenogenic activity of a Colombian strain of *Dunaliella salina*. The microalga accumulates high concentrations of b-carotene as an antioxidant photoprotector and glycerol as an ormoregulatory agent (El-Baky et al., 2004).

Flesh color of fish is the first quality evaluated by consumers, and is therefore an important component of quality relevant to market acceptance (Ünal Şengör et al., 2019). Therefore, the effects of *D. salina* on Red Tilapia flesh color were determined in this study. In addition, blood profiles (RBC, PCV, glucose and TP) (Mohammed, 2018 a) as indicative of health were determined. Therefore, the aims of the present study were to investigate growth performances, morphological and chemical characteristics of Red Tilapia upon dietary *D. salina* (DS) supplementation (33.0, 66.0 & 100.0%).

MATERIALS AND METHODS

SITE OF STUDY AND MANAGEMENT

The study was conducted according to procedures approved by the Ethics Committee of Animal Experimentation of King Faisal University, Saudi Arabi from March to June 2019. Fingerlings of Red tilapia fish were obtained from the Saudi fish farm. The fingerlings were allowed to adapt for two weeks, weighed using digital balance (Chyo Petit Balance MK-500C-Japan) (14.37±0.15 g) and distributed randomly to four groups. Fish were kept controlled under 12 cycle of light and 12 dark period. The controlled temperature and relative humidity during the experimental period were 27.0 ±3°C and 60.0 ±10%, respectively. Fish were fed control diet or diets supplemented with *D. salina* (33.0, 66.0 & 100.0) twice or three times a day for 14 wk. Ingredients (g/kg) and chemical composition (%) of control and *Dunaliella salina* (DS) formulated diets used during the

study were shown in Table (1 and 2). The recorded parameters of water quality during the study were 7.5-8.4 mg/L dissolved oxygen, 6.5-7.2 pH, 27-29 °C temperature, and <0.002 mg/L total ammonia.

GROWTH PERFORMANCE AND FEED EFFICIENCY

Body weight were recorded biweekly using digital balance (Chyo Petit Balance MK-500C-Japan). Fish were anesthetized using 0.1 g/l Tricainemethane sulfonate and dried before weighing. Body weight gain were determined using by subtracting final body weight from initial body weight. Tanks were cleaned before feeding using symphony system. Diets were offered three time per day at 7.0 and 11.0 am and 2.30 pm. Feed efficiency was calculated by dividing feed intake to body weight gain (Table 3 and 4).

CHEMICAL ANALYSIS OF DIETS AND FISH

Ingredients of diets and fish samples were dried at 70°C in air oven for constant weight. Both diets and fish samples were ground and analyzed for determination of dry matter (DM), organic matter (OM), crude protein (CP), crude fibers (CF) and ether extract (EE) components (Van keulen & Young, 1977) (Table 1 and 5).

BLOOD SAMPLE COLLECTION AND ANALYSIS

Blood samples were collected from the caudal vein of three fish of each treatment at the end of experimental period (Figure 1). The hematological parameters including (erythrocyte counts [RBC], total leucocyte counts [WBC], and packed cells volume [PCV]) were determined using a methodology of Hepler (1966). In addition, glucose and total protein concentrations were determined (Mohammed et al., 2018; Mohammed 2018 a, b).

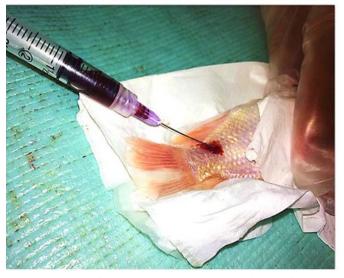


Figure 1: Blood sample collection throught caudal vein

COLOR MEASUREMENTS OF FISH MINCES

Color measurements of fish flesh minces were carried out using Hunter according to method of Young and Whittle

(1985). Mince color examination of L, a and b values are used to illustrate color differences between samples.

STATISTICAL ANALYSIS

Statistical analysis was done according to general linear model (GLM) of SAS program (2008). Differences between control and *D. salina* treated groups (33.0, 66.0 & 100.0%) were evaluated in growth performance, morphological and chemical body characteristics of fish, blood profiles by one-way ANOVA. Duncan Multiple Range Test was used to test the effect of treatments (Steel and Torrie, 1980). The data were presented as mean ± SEM. Level of significance was set at P < 0.05. Statistical model as follow:

Yij= μ + Ti + Eij

Where: Yij= the experimental observation ij, μ =the overall mean, Ti = the effect due to treatment i., Eij = the experimental error.

RESULTS

GROWTH PERFORMANCE AND FEED EFFICIENCY

Growth performance and feed efficiency due to feeding 33, 66, and 100% of D. salina to Red tilapia fish were presented in Tables (3-4). Results showed a significant decrease (p<0.05) in most of growth performance parameters of D. salina groups compared to control one, except survival rate, which was not differed (p>0.05) among groups. Furthermore, no significant difference (p<0.05) was obtained in the length-weight relationship, fish depth and conditional factor of 33, 66, and 100% D. salina groups compared to the control one (Tables 3-4). Supplementing 33, 66, and 100% of D. salina resulted in a significant decrease (p<0.05) in feed intake, feed efficiency compared to the control diet. Diet containing 33% of D. salina had no significant effect in protein productive value when compared with control diet versus 66 and 100% D. salina diets, which decreased the parameters. Productive value of energy was not differed (P > 0.05) among control and *D. salina* treated groups.

CHEMICAL COMPOSITION CHARACTERISTICS

Chemical composition characteristics of Red tilapia fish upon feeding 33, 66, and 100% of *D. salina* compared to control were presented in Table (5). In term of fish chemical component parameters, adding 33, 66, and 100% of *D. salina* resulted in no differences (p > 0.05) in values of dry matter, protein, ash and energy. However, values of fat were significantly increased when fish fed 66 and 100% of *D. salina* compared to the control diet.

Morphological Characteristics

Morphological characteristics due to feeding 33, 66, and 100% of *D. salina* to Red tilapia fish compared to control



Table 5: Chemical body composition of Red Tilapia fed diet supplemented with Dunaliella salina

Parameters	Control	DS 33%	DS 66%	DS 100%
Moisture, %	72.41±0.42a	72.33±0.36ª	72.57±1.01ª	72.21±1.03 ^a
Dry matter, %	27.59±0.36 ^a	27.67±1.01 ^a	27.43±1.03ª	27.79±0.54 ^a
Protein, %	15.28±0.07 ^a	15.48±1.00 ^a	14.51±1.15 ^a	14.26±0.63 ^a
Fat, %	7.53±0.19°	7.94±0.46 ^{bc}	8.46±0.55ab	8.90±0.54 ^a
Ash,%	4.78 ± 0.19^{a}	4.26±0.13	4.45±0.43 ^a	4.63±0.29 ^a
Energy, Calorie	$1.57 \pm 0.02a$	1.62±0.05 ^a	1.62±0.01 ^a	1.64±0.02 ^a
Energy, K Jole	6.53±0.09 ^a	6.73±0.22 ^a	6.71 ± 0.06^{a}	6.83±0.17 ^a

a,b,c; Values with different superscripts between DS (D. salina) and control groups significantly differed at P < 0.05.

Table 6: Morphological characteristics of Red Tilapia fed diet supplemented with Dunaliella salina

Parameters	Control	DS 33%	DS 66%	DS 100%
Body weight, g	27.06±5.65 ^a	$25.70 \pm 5.58^{\mathrm{ab}}$	24.39±4.29 ^b	21.8±3.22 ^c
Total length, cm	11.75±0.79 ^a	$11.56 \pm 0.77^{\mathrm{ab}}$	11.37±0.67 ^{bc}	11.11±0.51°
Standard length, cm	9.70±0.69ª	$9.48{\pm}0.67^{\mathrm{ab}}$	9.40±0.52 ^{bc}	9.16±0.46°
Body depth, cm	3.29±0.20 ^a	3.23±0.24 ^a	3.13±0.15 ^b	3.04±0.13°
Body thickness,	1.69±0.15 ^a	1.64 ± 0.19^{ab}	1.69±0.13 ^a	1.59 ± 0.10^{b}
Conditional factor	1.65±0.15	1.64±0.17	1.65±0.15	1.58±0.14

a,b,c; Values with different superscripts between DS (D. salina) and control groups significantly differed at P < 0.05.

Table 7: Blood parameters of Red Tilapia fed diet supplemented with Dunaliella salina

Parameters	Control	DS 33%	DS 66%	DS 100%
Red blood cell, 10 ⁶	2.81±0.37	2.72±0.28	2.75±0.32	2.78±0.29
Packed cell volume, %	29.50±2.78	28.73±2.51	29.133±3.83	29.67±3.21
Glucose, mg/100 ml	45.33±5.13	47.33±4.62	52.67±4.51	45.33±4.51
Total protein, g/100 ml	4.30±0.44	4.30±0.70	4.60±1.20	5.00±0.40

DS, D. salina

Table 8: Flesh coloration of Red Tilapia fed diet supplemented with Dunaliella salina

Parameters	Control	DS 33%	DS 66%	DS 100%
Lightness	58.08±2.26 ^a	$55.14 \pm 2.76^{\mathrm{ab}}$	56.91±2.45 ^a	53.66±2.27 ^b
Redness	6.31±1.07 ^b	8.92±1.27 ^a	8.66±1.65 ^a	8.55±1.04 ^a
Yellowness	16.42±0.21ab	16.25 ± 1.23^{ab}	17.09±0.57a	15.87±0.47 ^b

a,b,; Values with different superscripts between DS (D. salina) and control groups significantly differed at P < 0.05.

were presented in Table (6). The results showed that total length (cm), body depth (cm), and body thickness (cm) were significantly the lowest in 100% *D. salina* groups compared to the other groups, as well as conditional factor were found (P > 0.05) the lowest in 100% DS group.

BLOOD PROFILES

Blood profiles values (RBCs, Ht, glucose, TP) upon feeding 33, 66, and 100% of *D. salina* to Red tilapia fish compared to control were presented in Table (7). Results showed that adding 33, 66, and 100% of *D. salina* did not show any significant effect (p<0.05) in red blood cell values, hematocrit, total protein and glucose concentrations when compared to control diet.

COLOR MEASUREMENTS OF FISH MINCES

Fish minces coloration parameters upon feeding 33, 66, and 100% of *D. salina* to Red tilapia fish compared to control diet were presented in Table (8). Results showed that 100% *D. salina* diet caused a significant effect (p<0.05) in term of white flesh compared to control diet. There was a chronologically gradual in white color of fish minces. The highest was in the control group followed by 66, 33, and 100% groups, respectively. The diet containing 100% of *D. salina* caused a significant change in turning from whitish to yellowish flesh when compared with the control diet.

The need to continuous supplements of nutrients and feed inputs is required for the growth of aquaculture sector (Mustafa and Nakagawa 1995; Tacon et al., 2011) to meet the increase in human population. Several studies in the last decades were carried out using feed ingredients for farmed fish and crustaceans as protein source e.g. D. salina. Results of the current study demonstrated the effects of 33.0, 66.0 & 100% D. salina supplementation on Red Tilapia growth performances, feed efficiency, morphological and chemical body characteristics, flesh color and blood profiles (Table 3-8). The results indicated that replacement of fish meal with D. salina at low level (33.0%) resulted in comparable results with control diet concerning body weight gain, feed efficiency, morphological and chemical body characteristics and blood profiles. On the other hand, the high levels of D. salina (66.0 & 100) were mostly decreased the recorded parameters compared to control group except fat content and flesh color, which were significantly (P < 0.05) improved. The negative effects of 66.0 and 100.0% D. salina supplementation compared to control and 33.0% D. salina diets on growth performances and feed efficiency might be attributed to palatability problems (Walker and Berlinsky 2011) in addition to imbalances in diet components. Numbers of studies (Xu et al., 1993; Mustafa et al., 1995) were indicated improvement of body fat contents due to algae feeding as in 66.0 & 100.0% D. salina groups of this study. The survival rate and productive value of energy was not differed (P >0.05) among control and D. salina treated groups in this study. Algae of D. salina has high levels of beta-carotene, glycerol, protein and other fine chemicals (Gouveia et al., 2008; Ghasemi et al., 2011; Wichuk et al., 2014; Cuellar-Bermudez et al., 2015; Gong and Bassi, 2016). Chemical composition of D. salina algae indicated 8.47% moisture, 54.17% crude protein, 0.80% fiber, 11.42% total lipid, 18.47% ether extract and 6.67% ash (Mohammed 2018 b). The effects of partial replacement of fish meal protein by microalgae on body growth, feed intake, and body composition of Atlantic cod were investigated by Walker and Berlinsky (2011). They found no differences in survival and feed conversion ratios whereas feed intake and growth were significantly reduced in the algae-fed fish. In addition, feed intake improved in the 15% algae-fed fish compared to those fed the diet with 30% replacement. Alishahi et al. (2013) investigated the immunological responses of Heros severus fed diet supplemented with D. salina. They found that D. salina were positively affected the immunological parameters, which might be attributed to the comparable survival rate of control and D. salina groups in our study. The non-significant changes of fish survival rate between control and D. salina groups has been confirmed through blood profiles (RBC, PCV, glucose and TP) as indicative of fish health (Table 8) in this study.

Flesh color of fish is the first quality parameter evaluated by consumers, and is therefore an important component of quality relevant to market acceptance (Ünal Şengör et al., 2019). The significant improvement of flesh color due to 66.0 and 100.0% D. salina supplementation compared to control and 33.0% DS diets were obtained in this study, which might be attributed to pigment content of *D. salina*. Pigment contents of algae like, chlorophylls, carotenoids, phycobiliproteins, xanthophylls were confirmed (Roy and Ruma, 2014). Lipophilic pigments such as chlorophylls and carotenoids constitute 3-5 % of the dry algal biomass (Venkataraman and Becker, 1985).

CONCLUSION

Supplementation of D. salina could be promising of Red Tilapia growth performances, feed efficiency and body chemical characteristics at level 33.0% or low.

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CONFLICTS OF INTEREST

There is no conflict of interest of this article to declare.

AUTHORS CONTRIBUTION

All authors contributed equally.

REFERENCES

- Abdel-Daim MM, Farouk SM, Madkour FF, Azab SS (2015). Anti-inflammatory and immunomodulatory effects of Spirulina platensis in comparison to Dunaliella salina in acetic acid-induced rat experimental colitis. Immunopharmacol. Immunotoxicol. 37 (2):126- 139. https://doi.org/10.3109/0 8923973.2014.998368
- ·Alishahi M, Karamifar M, Mesbah M, Zare M (2013). Hemato-immunological responses of Heros severus fed diets supplemented with different levels of Dunaliella salina. Fish Physiol. Biochem. 40:57-65. https://doi.org/10.1007/ s10695-013-9823-5
- •Bitterman N, Melamed Y, Ben-Amotz A (1994). b-Carotene and CNS oxygen toxicity in rats. J. Appl. Physiol. 76:1073-1076. https://doi.org/10.1152/jappl.1994.76.3.1073
- Cuellar-Bermudez SP, Aguilar-Hernandez I, Cardenas Chavez DL, Ornelas-Soto N, Romero-Ogawa MA, ParraSaldivar R (2015). Extraction and purification of high value metabolites from microalgae: essential lipids, astaxanthin and phycobiliproteins. Microb. Biotechnol. 8: 190-209. https://doi.org/10.1111/1751-7915.12167
- •El-Baky A, El-Baz F, El-Baroty G (2004). Production of

- antioxidant by the green alga Dunaliella salina. Int. J. Agric. Biol. 6, 49–57.
- El-Bermawi N (2013). Effects of dietary Dunaliella salina extract and highly unsaturated fatty acids on the fecundity and lipid content of pond-reared *Penaeus japonicus* broodstock. Commun Agric. Appl. Biol. Sci. 78(4):111-4.
- Gallego-Cartagena E, Castillo-Ramírez M, Martínez-Burgos W (2019). Effect of stressful conditions on the carotenogenic activity of a Colombian strain of Dunaliella salina. Saud. J. Biol. Sci. 26 (7): 1325-1330. https://doi.org/10.1016/j.sjbs.2019.07.010
- Ghasemi Y, Rasoul-Amini S, Morowvat MH (2011). Algae for the production of SCP. In: Liong, M.T. (Ed.), Bioprocess Sciences and Technology. Nova Science Publishers Inc. 163–184.
- Gong M, Bassi A (2016). Carotenoids from microalgae: A review of recent developments. Biotech. Adv. 34: 1396-1412. https://doi.org/10.1016/j.biotechadv.2016.10.005
- Gouveia L, Batista AP, Sousa I, Raymundo A, Bandarra NM (2008). Microalgae in novel food products. In K. Papadoupoulos, Food Chemistry Research Developments, pp. 75-112. New York: Nova Science Publishers.
- Hemmingson JA, Falshaw R, Furneaux RH, Thompson K (2006). Structure and antiviral activity of the galactofucan sulfates extracts from Undaria pinnatifida (Phaeophyta).
 J. Appl. Phycol. 18(2):185-193. https://doi.org/10.1007/s10811-006-9096-9
- Hepler, O. E. (1966). Manual of clinical laboratory method. Springfield, IL: Thomas.
- Krinsky NI (1988). Membrane antioxidants. Ann NY Acad. Sci. 551: 17-32. https://doi.org/10.1111/j.1749-6632.1988. tb22317.x
- Lavy A, Naveh Y, Coleman R, Mokady S, Werman MJ (2003). Dietary Dunaliella bardawil, a beta-carotenerich alga, protects against acetic acid-induced small bowel inflammation in rats. Inflamm. Bowel. Dis. 9(6):372-379. https://doi.org/10.1097/00054725-200311000-00005
- Mohammed AA (2018 a). Ovarian Tissue Transplantation in Mice and Rats: Comparison of Ovaries Age. Pak. J. Zool. 50(2): 481-486. https://doi.org/10.17582/journal. pjz/2018.50.2.481.486
- •Mohammed AA (2018 b). Development of oocytes and preimplantation embryos of mice fed diet supplemented with dunaliella salina. Adv. Anim. Vet. Sci. 6(1): 33-39. https://doi.org/10.17582/journal.aavs/2018/6.1.33.39
- Mohammed AA, Al-Hozab A, Alshaheen T (2018). Effects of Diazepam and Xylazine on Changes of Blood Oxygen and Glucose Levels in Mice. Adv. Anim. Vet. Sci. 6 (3): 121-127. https://doi.org/10.17582/journal.aavs/2018/6.3.121.127
- •Mustafa MG and Nakagawa H (1995). A review: dietary benefits of algae as an additive in fish feed. Isr J Aquac-

- Bamidgeh. 47: 155-162.

 Mustafa MG, Wakamatsu S, Takeda T, Umino T, Nakagawa H (1995). Effect of algae meal as feed additive on growth, feed efficiency and body composition in red sea bream. Fisheries Sci. 61: 25-28. https://doi.org/10.2331/fishsci.61.25
- Roy SS and Ruma P (2014). Microalgae in Aquaculture: A Review with Special References to Nutritional Value and Fish Dietetics. Proc. Zool. Soc. DOI 10.1007/s12595-013-0089-9. https://doi.org/10.1007/s12595-013-0089-9
- Roy SS and Ruma P (2014). Microalgae in Aquaculture: A Review with Special Referencesto Nutritional Value and Fish Dietetics. Proc. Zool. Soc. DOI 10.1007/s12595-013-0089-9. https://doi.org/10.1007/s12595-013-0089-9
- Senosy W, Kassab AY, Mohammed AA (2017). Effects of feeding green microalgae on ovarian activity, reproductive hormones and metabolic parameters of Boer goats in arid subtropics. Theriogenol. 96: 16-22. https://doi.org/10.1016/j. theriogenology. 2017.03.019
- Steel RG, Torrie JH (1980). Principles and Procedures of Statistics, A Biometrical Approach (2ndEd.) Mc Grow-Hill Book Co., New York.
- Tacon, AGJ Hasan, MR, Metian M (2011). Demand and supply
 of feed ingredients for farmed fish and crustaceans: trends
 and prospects. FAO, Fisheries and Aquaculture Technical
 Paper, No. 564, FAO, Pp 87.
- •Ünal Şengör GF, Balaban M O, Topaloğlu B, Ayvaz Z, Ceylan Z, Doğruyol H (2019). Color assessment by different techniques of gilthead seabream (Sparus aurata) during cold storage. Food Sci Technol. 39(3): 696-703. https://doi.org/10.1590/fst.02018
- Van keulen J, Young BA (1977). Evaluation of acid insoluble ash as a natural marker in ruminant digestibility studies. J Anim Sci. 44 (2): 282–287. https://doi.org/10.2527/jas1977.442282x
- Venkataraman, LV, Becker, EW (1985). Biotechnology and utilization of algae—The Indian experience. Mysore: Central Food Technological Research Institute. 257.
- •Walker AB, Berlinsky DL (2011). Effects of partial replacement of fish meal protein by microalgae on growth, feed intake, and body composition of Atlantic cod. North American Journal of Aquaculture, 73, 76–83.
- •Wichuk K, Brynjólfsson S, Fu W (2014). Biotechnological production of value-added carotenoids from microalgae. Bioengineered. 95(9): 5269–5275.
- •Xu B, Yamasaki S, Hirata H (1993). Supplementary Ulva sp. var. meal level in diet of Japanese flounder, Paralichthys olivaceus. Aquaculture Science 41: 461-468.
- Young KW, Whittle KJ (1985). Colour Measurement of Fish Minces Using Hunter L, a, b Values. J. Sci. Food Agric.36: 383-392. https://doi.org/10.1002/jsfa.2740360511

