Research Article



Efficacy of Aerosol Vaccination of Adjuvantd Pigeon Pox Vaccine in Pigeon

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Abstract | The current study was conducted to evaluate the adjuvanted aerosol vaccination of pigeon pox vaccine (PPV) with Gel 01 in comparison with feather follicle non adjuvanted route of vaccination. In order to determine the best aerosol field dose of PPV, challenge was done for 3 groups of pigeon (group no 2, 3 and 4) aerosolly vaccinated with non adjuvanted prepared live attenuated Vero cell tissue culture adapted PPV different field dose (Log 10 ^{2,5,3,5}, and $^{4.5}$ TCID₅₀) the protection percent were 75%, 80% and 80% respectively as field dose of $10^{3.5}$ TCID₅₀ was the field dose of choice to be used with different concentrations of Gel 01 adjuvant. Four groups of pigeon were vaccinated with PPV; group (1) vaccinated via feather follicle route in addition to other 3 groups of pigeon (group 5, 6 and 7) vaccinated via aerosol route with different Gel 01 adjuvant (5%, 10% and 15% respectively). Serum samples were collected weekly for 6 weeks. The highest level of induced antibodies detected by virus neutralization test (VNT) and ELISA were (3.0, 2.0, 2.5 and 2.5 NI respectively) and (1.9, 1.5, 1.8 and 1.7 S/P respectively) at the 3rd week post vaccination. The level of antibodies reduced dramatically in pigeon groups vaccinated via aerosol route group (5, 6 and 7) reached (1.5, 1.75 and 2.0 NI respectively) and (1.2, 1.3 and 1.4 S/P respectively) at 6th week on contrast to group (1) vaccinated with feather follicle route has a slight decrease in antibody level reached 2.75 NI and 1.8 S/P at the 6th week post vaccination. Negative result for VNT and ELISA were found in control non vaccinated group all over the period of study. All vaccinated four pigeon groups and non vaccinated control group were challenged by inoculation of the virulent PPV after 3 weeks from vaccination showing protection percent of 95%, in group (1) vaccinated by feather follicle while it was 85%, 90% and 90% in groups vaccinated via aerosol route with different Gel 01 percent (5%, 10% and 15% respectively) showing that 10 % Gel 01 is the concentration of choice to be used with PPV via aerosol route. On contrast, the control non-vaccinated pigeons showed 0% of protection. This study recorded the efficacy of aerosol vaccination of tissue culture adjuvanted PPV in pigeon using Gel 01 comparison to usual routine vaccination with unadjuvanted PPV by feather follicle route to establish a more safe route of vaccination in valuable oriental and racing pigeons and easier route of vaccination of large pigeon flock with recommendation of booster vaccination after 3 weeks in order to maintain a high level of protection and more duration of immunity.

Keywords | Pigeon Pox, Aerosol, Gel 01, ELISA, Vaccine.

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Pigeon pox is a serious virus disease in pigeons especially in young pigeon as causing high percentage of mortality due to (diphtheritic form) fibrino-necrotic lesion in the mucous membrane of the upper respiratory tract and (cutaneous form) the development of discrete proliferative nodular skin lesions (Hemanth et al., 2014).

Pigeon pox disease is caused by pigeon pox virus (PPV) within family Poxviridae subfamily Chordopoxvirinaeand genus Avipoxvirus (Andraw, 2012). The economic importance of the disease caused by the losses of infected pigeons, also due to cutaneous diphtheritic or systemic form as the lesions around the eyes and mouth with ulcerated oral cavity making pigeons unable to drink or eat causing dehydration or starvation leading to death (Hemanth et al ., 2014).

Vaccination is the ideal method for protection of pigeons from pigeon pox infection PPV vaccine can be used against pigeon and fowl poxvirus infection and also (Simon and Morten, 2011).

In Egypt, first pigeon pox vaccine was prepared by adaptation of pigeon pox virus (Hungarian strain) on the chorioallantoic membrane (CAM) of 9-11 day-old embryonated chicken eggs (Helmy et al., 1967). But recently alternative PPV vaccine was prepared by using locally isolated strain attenuated, adapted and propagated on Vero cell line (Kafafy et al., 2018).

The using of feather follicle method as a method for bird vaccination may have disadvantage as the post vaccinal effect and tick may reduce the values of some viable racing and demestic pigeons, in addition to the great efforts in manually bird vaccination (Gottstein et al., 2004). So the using of PPV vaccine can be given by aerosol route for protection against a pathogenic strain of virus given cutaneously (Mockett et al., 1990) especially its know that the common mode of infection for pigeon pox is via respiratory tract (OIE 2018). The antigen delivery to mucosa enhances local and systemic immune responses. So it provides an advantageous for vaccination on contrast to antigen injection (Kang et al., 2004).

Vaccination via mucosa by spray is a rapid and preventive method of vaccination in outbreaks in endemic areas and it should be suitable for mass application even applied by spray or aerosole and effective after single application (de Geusa et al., 2011; Shakya and Nandakumar., 2012) mucosal vaccination through inhalation of antigen alone may cause a tolerance state by contacting to respiratory tract mucosa so using virus antigen causes low immunogenic re-

sponse than using a polymer based adjuvant Montanide™ Gel 01 ST (Gel 01) (gel particles of sodium polyacrylate in water) which trigger the immune response of mucosal vaccination and overcome the problems of tolerance (Tseng et al., 2009).

Recently, Gel 01 polymer adjuvant is used to formulate mucosal bivalent inactivated Newcastle and Avian Influenza (H9N2) administrated through spray route produce protective humoral immune and cell-mediated response as well as challenge (Heba M E et al., 2017).

The present study aimed to prepare adjuvanted PPV vaccine with Gel 01 and evaluate the efficacy of aerosol vaccination with the prepared vaccine in pigeons.

MATERIAL AND METHODS

VIRUSES

Vaccinal strain: Pigeon pox vaccine was prepared on Vero cell line supplied by Pox Department, Veterinary Serum and Vaccine Research Institute (VSVRI). It was used for preparation of adjuvant PPV with Gel 01 with virus titer of 10^{5.5}TCID₅₀/ml.

Virulent pigeon pox: Virulent PPV was kindly supplied by Pox Vaccine Department, (VSVRI), Abbasia, Cairo. It had a titer of 10⁵EID₅₀ / ml and used for challenge of experimentally vaccinated pigeons.

Adjuvant: Montanide™ Gel 01 ready to dilute polymeric adjuvant was used. It contains gel particles of sodium polyacrylate in water.

Susceptible pigeons: One hundred and sixty (160)susceptible pigeons of 45 day - old were used in this study. These pigeons were divided as follows: Table (1).

VACCINATION

Feather follicle vaccination: It was done for the attenuated tissue culture pigeon pox vaccine 10 $^{5.5}\mathrm{TCID}_{50}$ / ml using a field dose of 10 ^{2.5}TCID₅₀/ field dose using a feather follicle route acciording to OIE 2018 and Kaffafy et al. (2018).

Aerosol route: It was done for the attenuated tissue culture pigeon pox vaccine 10 $^{5.5}\rm{TCID}_{50}$ / ml with different field doses 10 $^{2.5,3.5}$ and $^{4.5}\rm{TCID}_{50}$ / field dose according to Mockett et al. (1990) as Twenty five pigeons were placed in a cardboard container (460 x 460 x 580 mm) and a fine aerosol created using a spray gun were used.



Table 1: Experimental design

Groups	No. of pigeons	Adjuvant	Route	TCID/dose
1	20	NA	by feather follicle	$10^{2.5} TCID_{50}$ / field dose
2	20	NA	by aerosol	10 ^{2.5} TCID ₅₀ / field dose
3	20	NA	by aerosol	$10^{3.5} TCID_{50}$ / field dose
4	20	NA	by aerosol	10 ^{4.5} TCID ₅₀ / field dose
5	20	Gel 01(5%)	by aerosol	10 ^{3.5} TCID ₅₀ / field dose
6	20	Gel 01(10%)	by aerosol	10 ^{3.5} TCID ₅₀ / field dose
7	20	Gel 01(15%)	by aerosol	10 ^{3.5} TCID ₅₀ / field dose
8	10	NA	NA	NA
9	10	Gel 01(10%)	by aerosol	3X

Group (1): pigeons were vaccinated with PPV vaccine by feather follicle route.

Group (2): pigeons were vaccinated with PPV vaccine by aerosol without using of Gel 01 (using of $10^{2.5}TCID_{50}$ / field dose)

Group (3): pigeons were vaccinated with PPV vaccine by aerosol without using of Gel 01 (using of $10^{3.5}TCID_{50}$ / field dose)

Group (4): pigeons were vaccinated with PPV vaccine by aerosol without using Gel 01 (using of 10 ^{4.5}TCID₅₀/ field dose)

Group (5): pigeons were vaccinated with PPV vaccine by aerosol with using Gel 01 (5 %).

Group (6): pigeons were vaccinated with PPV vaccine by aerosol with using Gel 01 (10%).

Group (7): pigeons were vaccinated with PPV vaccine by aerosol with using Gel 01 (15 %).

Group (8): were kept as negative control non vaccinated.

Group (9): Ten susceptible pigeons were vaccinated with 10x PPV vaccine by aerosol with using Gel 01 (10%). For safety test

Table 2: Mean NI, S/P ratio and challenge percent of vaccinated pigeons with different field doses via spray

Group no	Mean NI after 21 days	Mean S/P ratio After 21 days	No. of birds showing lesion post challenge in 20 birds			Protection percent	
			5dpc	7dpc	10dpc	(%)	
Group (2) 10 ^{2.5} TCID ₅₀ / field dose)	1.5	1.1	1	22	22	75%	
Group (3) 10 ^{3.5} TCID ₅₀ / field dose	1.75	1.3	0	2	2	80%	
Group (4) 10 ^{4.5} TCID ₅₀ / field dose)	2.00	1.3	0	3	1	80%	

Titer of choice is group no (3) 10 3.5TCID₅₀/ field dose

Neutralization Index (NI) calculated as Virus Titer (VT) minus Serum Virus Titer (SVT)

 $NI \ge (1.5)$ consider as positive results

SERUM

Pigeon pox hyper-immune sera: Hyper-immune serum against PPV was obtained from Pox Research Department VSVRI and used in Serum Neutralization Test as a positive control

Serum samples: Serum samples were collected from all birds weekly before and after vaccination and challenge for detection of antibody levels by Serum Neutralization Test.

VERO CELL CULTURE

Vero cell line were kindly supplied from Pox Research Department, VSVRI and used for SNT

EARLE'S MINIMUM ESSENTIAL MEDIUM (MEM).

It was obtained from Sigma Chemical Company, USA and used as growth medium containing 10 % Newborn calf serum or as maintance medium containing 2% Newborn calf serum to use for SNT.

EVALUATION OF THE ADJUVANTED GEL 01 PIGEON POX VACCINE (QUALITY CONTROL): Sterility: It was carried out according to OIE (2018), where the mixed adjuvanted PPV vaccines were inoculated separately into tubes of nutrient agar, Sabouraud agar and thioglycolate medium and mycoplasma medium too.



Table 3: Results of NI and SP ratio of sera collected from vaccinated pigeons.

	Group	(1)	Group	(3)	Group	(5)	Group	(6)	Group	(7)	Cont group	rol Negative p(8)
Weeks	NI	S/P	NI	S/P	NI	S/P	NI	S/P	NI	S/P	NI	S/P
Pre vaccination	0.50	0.4	0.50	0.50	0.50	0.3	0.50	0.3	0.50	0.5	0.50	0.3
1WPV	1.25± 0.64	0.7± 0.46	0.75± 0.37	0.6± 0.28	1.00± 0.34	0.7± 0.26	1.25± 0.43	0.8± 0.3	1.25± 0.4	0.6± 0.36	0.25	0.5
2WPV	2.25± 0.64	1.6± 0.46	1.50± 0.37	1.0± 0.28	1.50± 0.34	1.1± 0.26	1.75± 0.43	1.3± 0.3	2.00± 0.4	1.4± 0.36	0.25	0.4
3WPV	3.00± 0.64	1.9± 0.46	1.75± 0.37	1.4± 0.28	2.00± 0.34	1.5± 0.26	2.50± 0.43	1.8± 0.3	2.50± 0.4	1.7± 0.36	0.25	0.5
4WPV	2.75± 0.64	1.7± 0.46	1.75± 0.37	1.3± 0.28	1.75± 0.34	1.2± 0.26	2.25± 0.43	1.5± 0.3	2.25± 0.4	1.4± 0.36	0.25	0.6
5WPV	2.75± 0.64	1.9± 0.46	1.50± 0.37	1.1± 0.28	1.75± 0.34	1.1± 0.26	2.00± 0.43	1.4± 0.3	2.00± 0.4	1.3± 0.36	0.25	0.5
6WPV	2.75± 0.64	1.8± 0.46	1.50± 0.37	1.1± 0.28	1.50± 0.34	1.2± 0.26	1.75± 0.43	1.3± 0.3	2.00± 0.4	1.4± 0.36	0.25	0.4

Group (1): susceptible pigeons were vaccinated with PPV vaccine feather follicle route

Group (3): susceptible pigeons were vaccinated with PPV vaccine by aerosol with 10 3.5 TCID₅₀/ field dose)

Group (5): susceptible pigeons were vaccinated with PPV vaccine by aerosol with using of Gel 01 (5 %).

Group (6): susceptible pigeons were vaccinated with PPV vaccine by aerosol with using of Gel 01 (10%).

Group (7): susceptible pigeons were vaccinated with PPV vaccine by aerosol with using of Gel 01 (15 %).

Table 4: Protection of vaccinated and control chicks against virulent fowl pox virus

Challenge Time post	Birds group	No. of challenged pigeons/group	· ·			Protection percent (%)	
vaccination			5dpc	7dpc	10dpc		
1 month	Group (1)	20	0	0	1	95%	
	Group (3)	20	0	2	2	80%	
	Group (5)	20	0	2	1	85%	
	Group (6)	20	5	1	1	90%	
	Group (7)	20	0	1	1	90%	
	Group (8)	10	2	3	5	0%	

Group (1): twenty susceptible pigeons were vaccinated with PPV vaccine feather follicle route

Group (3): Twenty susceptible pigeons were vaccinated with PPV vaccine by aerosol without using of Gel 01

Group (5): Twenty susceptible pigeons were vaccinated with PPV vaccine by aerosol with using of Gel 01 (5 %).

Group (6): Twenty susceptible pigeons were vaccinated with PPV vaccine by aerosol with using of Gel 01 (10%).

Group (7): Twenty susceptible pigeons were vaccinated with PPV vaccine by aerosol with using of Gel 01 (15 %).

Group (8): Ten susceptible pigeons were kept as control non vaccinated.

Safety: Ten susceptible pigeons were vaccinated with 10 x field dose of adjuvanted Gel 01 PPV vaccine using aerosol mode according to OIE (2018).

Potency: Challenge test was applied by inoculation of the virulent pigeon pox virus by feather follicle route in vaccinated and susceptible control pigeons at 3 weeks post vaccination. All birds were subjected to daily observation of gross lesions and collection of serum samples according to OIE (2018).

SEROLOGICAL ASSAY

Serum neutralization test (SNT): Serum neutralization test (SNT) was conducted according to Kaffafy et al. (2018) and Amina and Christine (2020) for the detection of antibody levels after vaccination of different group \and results calculated according to Reed and Muench (1938).

Indirect enzyme linked immunosorbent assay (ELISA): It was carried out according to Kaffay et al. (2015)

DETERMINTION OF PPV VACCINE FIELD DOSE VIA AERSOL SPRAY

The results of mean NI and S/P ratio as well as challenge for defferent PPV vaccine field doses administrated via spray shown in Table (2) refer Protective level of antibody represented by NI and S/P ratio in all vaccinated groups while challenging the birds 3 weeks post vacciation with virulent PPV revealed that (5) birds showed post challenging pox lesions and the other 15 birds remained healthy without local or generalized post challenging lesion with protection percent of 75% in group (2) received 10 ^{2.5}TCID₅₀/ field dose. While the protective percent against challenge were 80% in both group (3) received 10 3.5TCID₅₀/ field dose and group (4) received 10 4.5 TCID₅₀/ field dose as (4) pigeon showed symptoms and the other 16 birds remained healthy without local or generalized post challenging. Table (2).

QUALITY CONTROL OF THE PREPARED PPV VACCINE

Sterility test: Bacterial culture of prepared adjuvanted PPV vaccine with gel 01 were proved to be free from any bacterial and fungal contamination.

Safety test: Using of 10 times of the recommended dose of adjuvanted PPV vaccine with gel 01 10% proved that the vaccine was safe to be used in pigeons. Where the vaccinated birds did not show any undesirable symptoms refer to the adjuvant or the vaccine virus.

Serum Neutralization test and indirect ELISA: Serum neutralization test as well as Indirect ELISA were carried on the serum samples collected from vaccinated pigeons via spray route without and with different concentrations of adjuvanted Gel 01 (groups no 3, 5, 6 and 7) (0%, 5%, 10% and 15% respectively) PPV vaccines and feather follicle method (group no 1). The results were expressed as neutralization index (NI) and (SP) ratio. The result which presented in Table (3) Figure (1) revealed that, the level of neutralizing antibodies in all vaccinated Pigeons serum samples as NI values were above 1.5 NI and S/P values above 1 after 2 weeks of vaccination and the highest antibody titers appeared at 3 weeks post vaccination in all groups but higher in group (1) as NI=3 and S/P=1.9 than other groups followed by decrease in antibody level after 3weeks post vaccination which is much more in groups (3, 5, 6 and 7) than group (1). Table (3) and Figure (1).

Result of challenge test in pigeons with virulent pigeon pox virus: Table (4) showed that after 3 weeks post vaccination the protection rate against PPV in group (1) vaccinated by feather follicle was 95% in compared with 80%, 85%, 90% and 90% in birds vaccinated via aerosol mode using different percent of Gel 01 0%, 5%, 10% and 15% respectively while in control group it was 0%. Table (4).

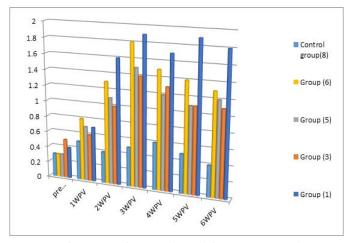


Figure 1: SP ratio of sera collected from vaccinated pigeons.

DISCUSSION

Pigeon pox disease is one of most important viral disease affecting pigeons due to its economic impact on pigeon rearing systems as it causes high fatality rate especially in young birds due to respiratory diphteric form, (Soad et al., 2007; Abdallah and Ola, 2013; OIE, 2018) even it causes cutaneous distortions in valuable oriental racing pigeons and domestic pigeon when infected by skin form (Sudhakara and Sivajothi, 2018). Vaccination is the only effective mean for controlling and preventing pigeon pox disease in pigeon (Amina and Christine, 2020).

Through the present study an innovation of new vaccination route for PPV via aerosol was performed and evaluated through using of adjuvanted Gel 01 PPV aerosol vaccine in comparison to the traditional feather follicle vaccination route in pigeon regarding to that one of most important route for PPV infection is via respiratory inhalation mode (OIE, 2018) in order to establishment an effective rapid and less overwrought method for vaccination of large pigeon flock farm and a safe vaccination way without post vaccinal reaction in oriental viable high price racing pigeon. Tissue culture propagated pigeon pox vaccine on Vero cell line with virus titer of 10 5.5 TCID₅₀/ ml was used in this study for aerosol vaccination (Aboul Soud et al., 2018) and (Kaffay et al., 2018), In order to determine the effective field dose 3 susceptible pigeon groups (group 2. 3.and 4) received different field doses of PPV via aerosol without adjuvant (10 $^{2.5}\mathrm{TCID}_{50}$, 10 $^{3.5}\mathrm{TCID}_{50}$ and 10 $^{4.5}\mathrm{TCID}_{50}/$ field dose) followed by antibody detection by (VNT and indirect ELISA) and challenging with virulent PPV at 21 post vaccination the result represented in Table (2) showed that group (3) received 10 ³⁵TCID₅₀/ field dose is the field dose of choice with NI 1.75 of SP 1.3 with protection percent of 80% as group 2 showing lower NI

1.5, SP 1.1 and protection percent of 75% while group 4 received higher field dose showing same protection percent of 80% with non-significant increased NI 2.0 and SP 1.3 similar results obtained by Nagy et al. (1990) when they determine the field dose of fowl pox vaccine (FPV) to be used via spray while it was 10 °TCID₅₀/ ml for 100 bird.

The humeral immune response (antibody response) to vaccine is an indirect measure of the immunogenicity of the vaccine and accordingly the immune status of vaccinated pigeons groups vaccinated with PPV via aerosol with different concentrations of Gel 01 (0%, 5%, 10 % and 15%) groups no (3, 5, 6 and 7 respectively) and group no (1) pigeons vaccinated with PPV vaccine through feather follicle was estimated and compared by Serum neutralization test (SNT) and ELISA.

As the observed results in Table (3) showed that protective level of antibody titer in group no 1, 3, 5, 6, and 7 expressed as NI = (2.25, 1.5, 1.75 and 2.0 –S/P values = 1.6, 1, 1.1, 1.3 and 1.4 respectively) appeared 2 weeks post vaccination while the highest level of antibody titer were at 3rd week post vaccination expressed as NI = 3.0,, 1.75, 2.0, 2.5 and 2.5 S/P values =1.9,, 1.4, 1.5, 1.8 and 1.7 respectively. These result is in accordance with (Kaffafy et al., 2018) and (Amina and Chrestine, 2020), they found that the level of vaccinated pigeon with egg adapted and tissue culture adapted PPV appeared 2 weeks post vaccination reaching its peak at 3 weeks post vaccination also (Nagy et al., 1990; Mockett.et al., 1990) when they used spray mode for FPV vaccine in chicksa compared with different routes of administrated.

From the previous data it was found that Gel 01 best concentration to be added to PPV vaccine used via aerosol was 10% as using 5% Gel 01 showed humeral immune response less than obtained by 10% Gel 01 while using higher concentration of Gel 01 showing non-significant increase iin the humeral immune response these results were in accordance with (Heba et al., 2017) who used 10 % Gel 01 for Preparation of mucosal nanoparticles inactivated vaccine for Newcastle disease and H9N2 AI viruses.

It was noticed that higher antibody level in group no (1) (vaccinated via feather follicle) than the other groups vaccinated with aerosol method. Also the dramatically decrease of the level of immune response in all groups vaccinated via aerosol route than group (1) from week 4 till week 6 post vaccination which is also reported by (Heba et al., 2017) who used adjuvanted spray mucosal vaccination by Gel 01 inactivated influenza H9N2 vaccine and explained the effect of adjuvanted aerosol mucosal vaccination on innate immunity rather than adaptive immunity which indicated urgent boster dose 4 weeks post vaccination (Joo et al.,

2011).

Challenge is considered the master test to evaluate and assist the immunizing capacity of the vaccine against pox infection, so all vaccinated groups were tested by challenging the immunity of the vaccinated pigeons against the virulent pigeon pox virus. The results represented in Table (4) showed that protection percent in group no (1) vaccinated with feather follicle route was 95% These results were in accordance with (Kaffafy et al., 2018) and (Amina and Chrestine, 2020) they found that protection percent for tissue culture adapted PP were 90% and 95% respectively when they compared with protection percent of PPV adapted on embryonated chicken egg (ECE) 100%, The protection percent in pigeon groups (no 3, 5, 6 and 7) vaccinated via aerosol with different Gel 01 percent (0%, 5%, 10% and 15%) were 80%, 85% 90% and 90% respectively as groups received PPV with 10 % and 15% Gel 01 gave the higher and equal protection percent 90% than other groups similar results obtained by (Nagy et al., 1990) when they vaccinated chicks with fowl pox (FP) vaccine via aerosol also (Mockett et al., 1990) found that no significant difference in protection percent in groups of chickens vaccinated with FPV via spray route and chickens vaccinated with wing web method as they mentioned that the experiment showed that aerosol route is a very effective route for FP vaccination. As a number of avian viruses enter their host by the respiratory route (infectious laryngotracheitis, avian influenza, infectious bronchitis and Newcastle disease viruses). A fowl pox given by the aerosol route would be expected to give rise to local immunity and this may be a very important factor in limiting spread of the avian viruses. Therefore, the aerosol route remains a practical alternative to the wing web route.

The increased level of humeral immune response and protective percent of challenge by adding 10 % Gel 01 to the PPV vaccine via aerosol which is higher than that of using PPV vaccine via aerosol without adjuvant can be explained on the basis of immune enhancing effects of Montanide adjuvants on vaccinated birds immune response which increase the efficacy of mucosal vaccination against avian infectious diseases (Shakya. and Nandakumar, 2012) as Gel 01 can enhance the efficacy of spray delivery of viral vaccines. This regarding to the muco adhesive nature of the polymer compounds which increase contact time with respiratory tract mucosa giving the chance to the innate immune response to take their role in protecting immune response (Adams et al., 2014). Montanide gel 01 induces strong infiltration of monocytes and macrophages, which enhances phagocytosis of the antigen complex with the polymer simultaneously increase the antigen-presenting cells activity and the innate immune response that trigger the acquired immune system to induce a highly specific

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immune response, so development of adjuvanted Gel 01 mucosal delivered vaccines may induce superior protection against viruses that infect via mucosal surfaces by stimulating the cellular immunity (Jang et al., 2011).

From the obtained results, the present study proved the capability of using PPV vaccine via aerosol route with Gel 01 adjuvant as mucosal vaccine with recommendation of poster 2^{nd} dose after 3 weeks post vaccination.

CONCLUSION

This study recorded the efficacy of aerosol vaccination of tissue culture adjuvanted PPV vaacine in pigeon using Gel 01 in comparison to the usual routine vaccination with non adjuvanted PPV vaccine by feather follicle route to establish a more safe route of vaccination in valuable oriental and racing pigeons and easier route of vaccination of large pigeon flock with recommendation of poster vaccination after 3 weeks of vaccination in order to maintain a high level of protection and more long duration of immunity.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest for this publication.

AUTHORS CONTRIBUTION

All authors contributed equally to study design, sampling, Methodology, interpretation of results, and writing of the manuscript.

REFERENCES

- •Aboul Soud AE, Olfat E Nakhla, Namaa A Mohamed, El Soally SA, Ayatollah II (2018) :Molecular and Antigenic Characterization of Pigeon Pox Virus Isolated in 2017. International Journal of Innovative Research & Development (Ijird), 2018: 7 (4): 219-231. https://doi.org/10.24940/ijird/2018/v7/i4/APR18091
- Abdallah FM, Ola FH (2013). Detection and molecular characterization of avipoxviruses isolated from different avian species in Egypt. Virus Genes (2013) 46:63–70 https://doi.org/10.1007/s11262-012-0821-y
- Adams JR, Haughney SL, Mallapragada SK (2014). Effective polymer adjuvants for sustained delivery of protein subunit vaccines. Acta Biomater.14:104-114. https://doi. org/10.1016/j.actbio.2014.11.050
- Amina AR, Christine AM (2020): Comparitive evaluation on the efficacy of emproynated chicken egg adapted and tissue

- culture pigeon pox vaccines against the local isolate strain. J. Appl. Vet. Sci. 5 (2): 87 93. https://doi.org/10.21608/javs.2020.85605
- Andraw K (2012). Virus taxonomy: ninth report of the international committee on the taxonomy of viruses. The International Committee on Taxonomy of International Union of Microbiological Societies. pp.298.
- De Geusa ED, van Haarlema DA, Poetria ON, de Wit JJ, Vervelde L (2011). A lack of antibody formation against inactivated influenza virus after aerosol vaccination in presence or absence of adjuvantia. Vet. Immunol. Immunopathol., 143: 143-147. https://doi.org/10.1016/j.vetimm.2011.05.023
- Gottstein ZG, Horvatek DE, CAJAVEC S, Mazija H (2004). Safety and immunogenicity of pigeon pox virus vaccine applied to newly hatched chickens by means of nebulization, In Book of Abstracts XXII World Poultry Congress, Istanbul, Turkey.
- Heba ME, Mohamed SM, Hussein AH (2017). Preparation of mucosal nanoparticles and polymer-based inactivated vaccine for Newcastle disease and H9N2 AI viruses. Vet. World. 10: 187-193. https://doi.org/10.14202/vetworld.2017.187-193
- Helmy N Ahmed, H El Sabbagh, Zahran G (1967). Studies on pigeon pox vaccine in the U.A.R. Preparation and Application of Desiccated chorioallantoic membrane vaccine. J. Egyptian Vet. Med. Assoc, Volxxvii:85-90.
- Hemanth I, Amaravathi P, Sasidhar BN, Anand KA, Sailaja N (2014). An out-break of cutaneous form of avian pox in pigeons (columbia livia). Int. J. Sci. Environ. Technol. 3(4): 1484 – 1488.
- Jang SI, Lillehoj HS, Lee SH, Lee KW, Lillehoj EP, Bertrand F, Dupuis L, Deville S (2011). Montanide™ IMS 1313 N VG PR nanoparticle adjuvant enhances antigen-specific immune responses to profilin following mucosal vaccination against Eimeria acervulina. Vet. Parasitol., 182: 163-170. https://doi.org/10.1016/j.vetpar.2011.05.019
- •Joo HM, He Y, Sundararajan A, Huan L, Sangster MY (2011). Quantitative analysis of influenza virus-specific B cell memory generated by different routes of inactivated virus vaccination. Vaccine. 28(10): 2186-2194. https://doi.org/10.1016/j.vaccine.2009.12.058
- Kafafy MH, Ahmed HA, Ali AS, Mohammed A Ismaeil, Manal A (2015). Isolation and Characterization of Pigeon Pox virus in Egypt. Zagazig University (virology) PHD faculty of veterinary medicine.
- Kafafy MH, Aboul- Souad EA, Olfat EN, Christine AM, Nermin ME (2018). Prelimery study on prepration and evalution of a local isolate tissue culture propagated pigeon pox vaccine. VMJG Vol (46): 44-50.
- Kang SM, Guo L, Yao Q, Skountzou I, Compans RW (2004):
 Intranasal immunization with inactivated influenza virus enhances immune responses to coadministered simianhuman immunodeficiency virus-like particle antigens.
 J. Virol., 78: 9624-9632. https://doi.org/10.1128/JVI.78.18.9624-9632.2004
- Mockett PM, Deuter A, Debra Southee J (1990). Fowlpox vaccination:Routes of inoculation and pathological effects, Avian Pathol. 19(4): 613-625. https://doi. org/10.1080/03079459008418717
- Nagy EA, Maeda AD, Krell PJ, Derbysshire JB (1990). Vaccination of 1-day-old chicks with fowl pox virus by aersol, drinking water or cutaneous routes. Avian Dis. (34): 677-682. https://doi.org/10.2307/1591263
- Office International Des Epizootic (OIE) (2018). Recommended



virulent strains of pigeon pox viruses Zag. Vet. J. 35(1): 8-13.

- Diagnostic Technique and Requirements for Biological Poducts, Volume 1 (Chapter 3.3.10. pp 906-913).
- Reed LJ, Munech H (1938). A simple method of estimating fifty percent end points. Am. J. Hyg,., 27:493-497. https:// doi.org/10.1093/oxfordjournals.aje.a118408
- •Shakya AK, Nandakumar KS (2012). Applications of Polymeric Adjuvants in Studying Autoimmune Responses and Vaccination against Infectious Diseases. J R Society Interface 10: 20120536. Medical Inflammation Research, Department of Medical Biochemistry and Biophysics, Karolinska Institute, Stockholm, Sweden. https://doi. org/10.1098/rsif.2012.0536
- Simon CW, Morten T (2011). Avipox viruses infection biology and their use as vaccine vectors». Viro. J., 8: 49. https://doi.

- org/10.1186/1743-422X-8-49
- Soad MS, Abd El-Razik BA, Amal F, Amira AE, Micheal A (2007). Antigenic comparative studies between vaccinal and
- Sudhakara RB, Sivajothi S (2018). Therapeutic management of cutaneous form of pox in pigeons with azithromycin. Int. J. Avian Wildlife Biol. 3(3):136–137. https://doi. org/10.15406/ijawb.2018.03.00073
- •Tseng LP, Chiou CJ, Chen CC, Deng MC, Chung TW, Huang YY, Liu DZ (2009). Effect of lipopolysaccharide on intranasal administration of liposomal Newcastle disease virus vaccine to SPF chickens. Vet. Immunol. Immunopathol., 3(4): 285-289. https://doi.org/10.1016/j.vetimm.2009.04.009

